

Supplementary Materials for  
**mTORC1 controls lysosomal Ca<sup>2+</sup> release through the two-pore channel  
TPC2**

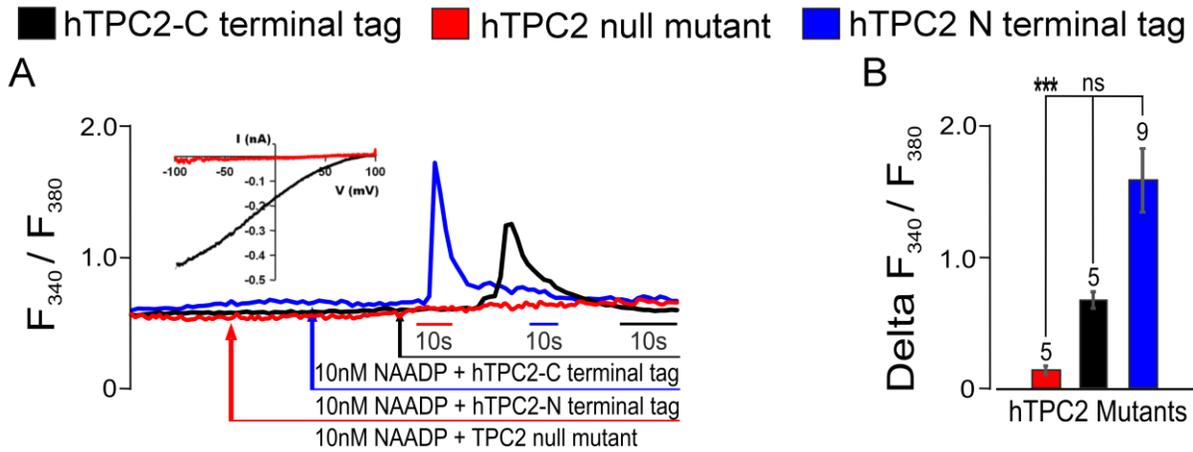
Oluseye A. Ogunbayo, Jingxian Duan, Jian Xiong, Qiaochu Wang, Xinghua Feng,  
Jianjie Ma, Michael X. Zhu, A. Mark Evans\*

\*Corresponding author. Email: mark.evans@ed.ac.uk

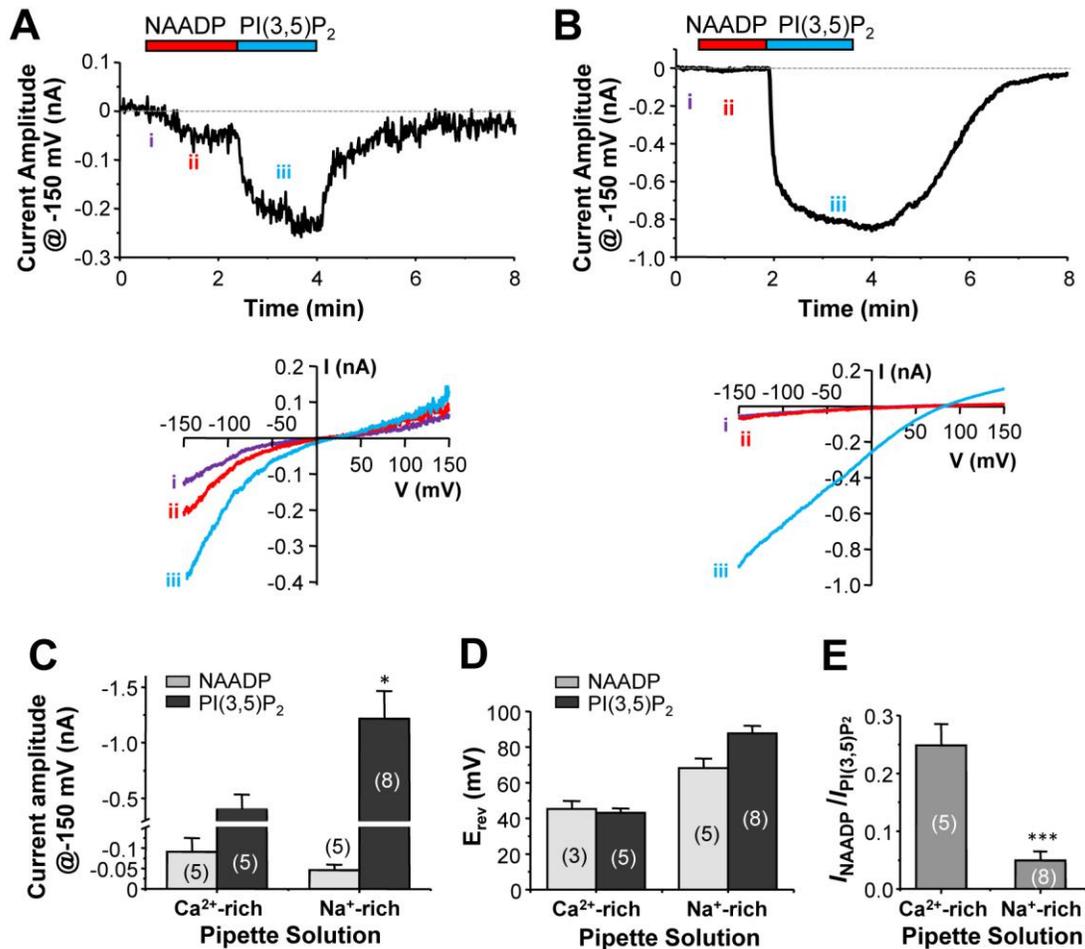
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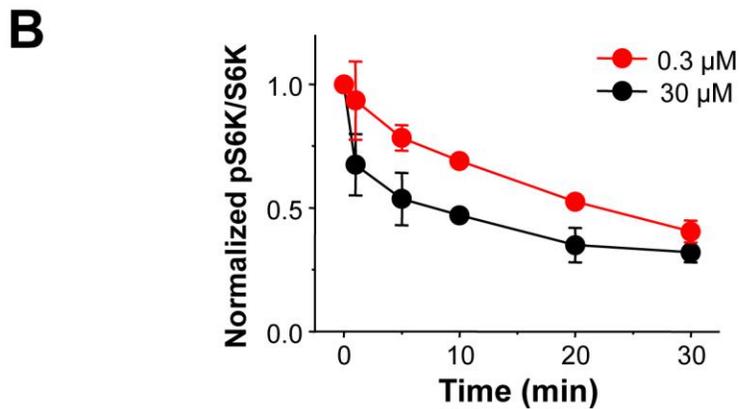
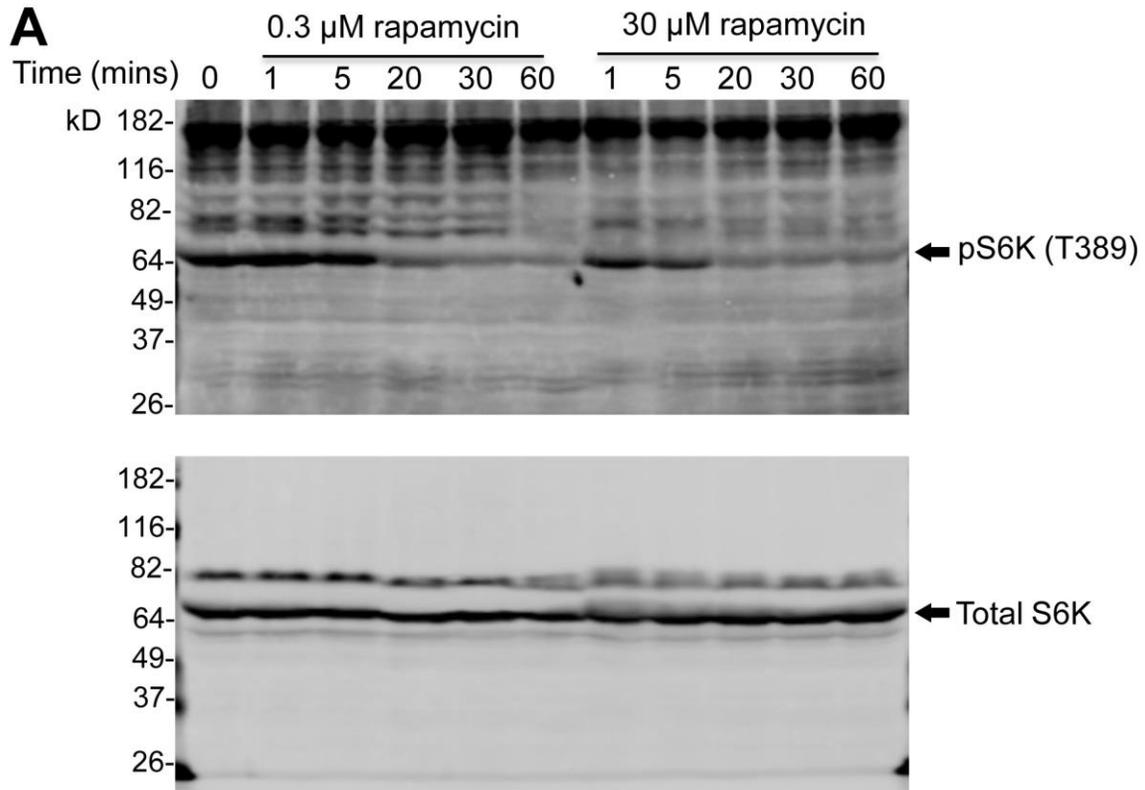
- Fig. S1. Blind experiments on active and null hTPC2 constructs demonstrate robustness of intracellular dialysis technique.
- Fig. S2. Na<sup>+</sup> and Ca<sup>2+</sup> currents mediated by endolysosomal TPC2 in response to NAADP and PI(3,5)P<sub>2</sub>.
- Fig. S3. High and low concentrations of rapamycin suppress mTORC1 activities in HEK293 cells at different rates.
- Fig. S4. Torin-2 induces increases in intracellular Ca<sup>2+</sup> in HEK293 cells stably overexpressing hTPC2 and in rat pulmonary arterial myocytes.
- Fig. S5. Torin-1 induces low-magnitude, sustained increases in intracellular Ca<sup>2+</sup> in HEK293 cells stably overexpressing hTPC2 and in rat pulmonary arterial myocytes.



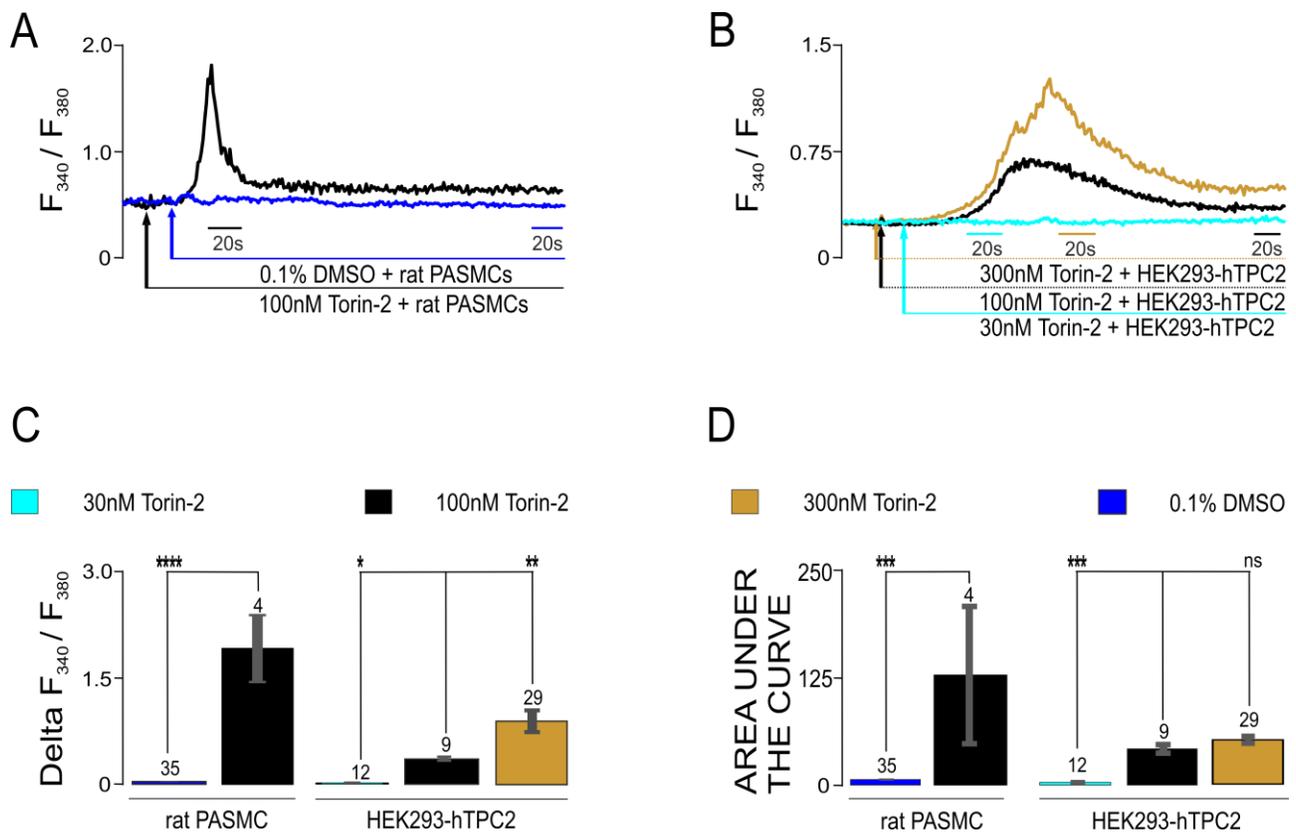
**Fig. S1. Blind experiments on active and null hTPC2 constructs demonstrate robustness of intracellular dialysis technique.** **A**, Records of  $F_{340} / F_{380}$  ratio against time during intracellular dialysis of 10 nmol/L NAADP into HEK 293 cells expressing either wild type hTPC2 with a C terminal EGFP tag (black), hTPC2 with an N terminal EGFP tag (blue) or an hTPC2 null mutant but lacking a  $PI(3,5)P_2$ -induced  $Na^+$  current (red); arrows indicate the beginning of intracellular dialysis upon entering the whole-cell configuration. Inset shows whole-endolysosomal current evoked by 1  $\mu$ mol / L  $PI(3,5)P_2$  recorded from enlarged vacuoles isolated from HEK 293 cells that expressed C-terminal EGFP-tagged hTPC2 (black) or the hTPC2 null mutant (red). **B**, bar chart showing mean  $\pm$  SEM for each construct tested. Number of experiments (n) indicated above bars \*\*\* $P < 0.001$ .



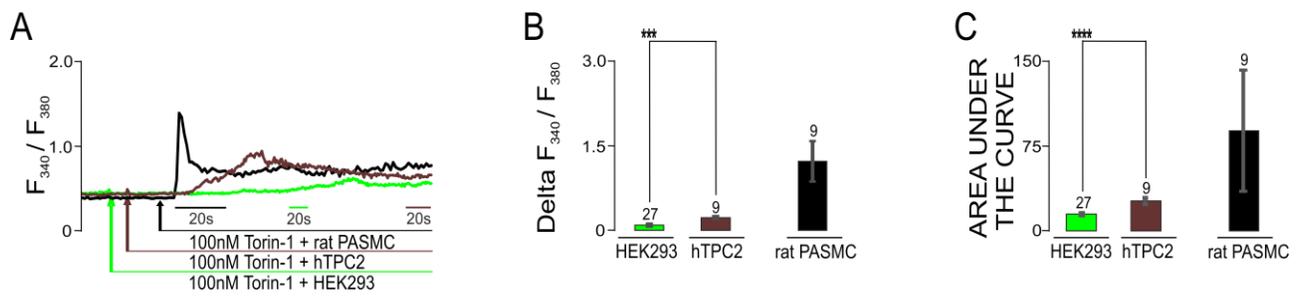
**Fig. S2. Na<sup>+</sup> and Ca<sup>2+</sup> currents mediated by endolysosomal TPC2 in response to NAADP and PI(3,5)P<sub>2</sub>.** **A** and **B**, representative traces of currents at -150 mV (*upper traces*) and current-voltage (I-V) curves at the indicated time points (*lower traces*). For **A**, the pipette solution was Ca<sup>2+</sup>-rich (Na<sup>+</sup>-free) and contained (in mmol / L) 70 K-methanesulfonate (MSA), 60 Ca-MSA, 1 MgCl<sub>2</sub>, 10 Hepes (pH adjusted with MSA to 4.6 and mannitol used to adjust osmolarity); the bath solution contained (in mmol / L) 130 K-MSA, 0.2 mM Ca-MSA, 10 Hepes (pH adjusted with KOH to 7.2). For **B**, the pipette solution was Na<sup>+</sup>-rich and contained (in mmol / L) 145 NaCl, 5 KCl, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub>, 10 Hepes, 10 MES, 10 glucose (pH adjusted with NaOH to 4.6); the bath solution contained (in mmol / L) 140 K-gluconate, 4 NaCl, 1 EGTA, 2 MgCl<sub>2</sub>, 0.39 CaCl<sub>2</sub>, 20 Hepes (pH adjusted with KOH to 7.2; free [Ca<sup>2+</sup>] = 100 nM). NAADP (100 nmol / L) and PI(3,5)P<sub>2</sub> (1 μmol / L) were applied by perfusion to the cytoplasmic side of the endolysosomal vacuole as indicated by the bars above the current trace. **C-E**, bar charts show the mean ± SEM for the peak of current amplitude at -150 mV (**C**) and reversal potential (E<sub>rev</sub>) of currents (**D**) evoked by 100 nmol / L NAADP and 1 μmol / L PI(3,5)P<sub>2</sub>, as well as ratio of peak current evoked by NAADP to that by PI(3,5)P<sub>2</sub> (**E**) in the Ca<sup>2+</sup>-rich or Na<sup>+</sup>-rich pipette solution. Values were determined after subtracting basal current. The numbers of vacuoles recorded are indicated in parentheses. For some vacuoles, the E<sub>rev</sub> was indiscernible because the evoked current was too small. \**P* < 0.05, \*\*\**P* < 0.001 vs. Ca<sup>2+</sup>-rich by unpaired *t* test.



**Fig. S3. High and low concentrations of rapamycin suppress mTORC1 activities in HEK293 cells at different rates.** **A**, Upper panel shows full immune-blot of phospho-S6K (pS6K) versus time as an index of mTORC1 activity for HEK 293 cells treated with either 0.3 or 30  $\mu\text{mol} / \text{L}$  rapamycin for the time periods indicated; at each time point cells were lysed immediately. Lower panel shows full immunoblot for total S6K at each time point. **B**, summary data for pS6K / total S6K ratios (normalized to untreated cells) compared to time. Each data point represents results from 2 to 4 experiments expressed as mean  $\pm$  SD or range.



**Fig. S4. Torin-2 induces increases in intracellular  $Ca^{2+}$  in HEK293 cells stably overexpressing hTPC2 and in rat pulmonary arterial myocytes.** **A**, Records of  $F_{340} / F_{380}$  ratio against time during extracellular application of 100 nmol / L torin-2 (black) and vesicle (0.1% DMSO, blue) on to a rat pulmonary arterial myocyte. **B**, Records of  $F_{340} / F_{380}$  ratio against time during extracellular application of 30 (cyan), 100 (black) and 300 (brown) nmol / L torin-2 on to HEK 293 cells stably overexpressing hTPC2. **C** and **D**, bar charts showing mean  $\pm$  SEM for each cell type and treatment for the peak change in Fura-2 fluorescence ratio (Delta  $F_{340} / F_{380}$ ; C) and area under the curve (D). Number of cells (n) indicated above bars. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ .



**Fig. S5. Torin-1 induces low-magnitude, sustained increases in intracellular  $Ca^{2+}$  in HEK293 cells stably overexpressing hTPC2 and in rat pulmonary arterial myocytes.** **A**, Records of  $F_{340} / F_{380}$  ratio against time during extracellular application of 100 nmol / L torin-1 on to a wild type HEK 293 cell (green), a HEK 293 cell stably overexpressing hTPC2 (brown) and a pulmonary arterial myocyte (black). Note, the fluorescence ratio did not return to baseline at the end of the recording. **B**, bar charts showing mean  $\pm$  SEM for each cell type for the peak change in Fura-2 fluorescence ratio (Delta  $F_{340} / F_{380}$ ; B) and area under the curve (C). Number of cells (n) indicated above bars. \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ .