

Supplementary Materials for

Thrombospondin-1 promotes matrix homeostasis by interacting with collagen and lysyl oxidase precursors and collagen cross-linking sites

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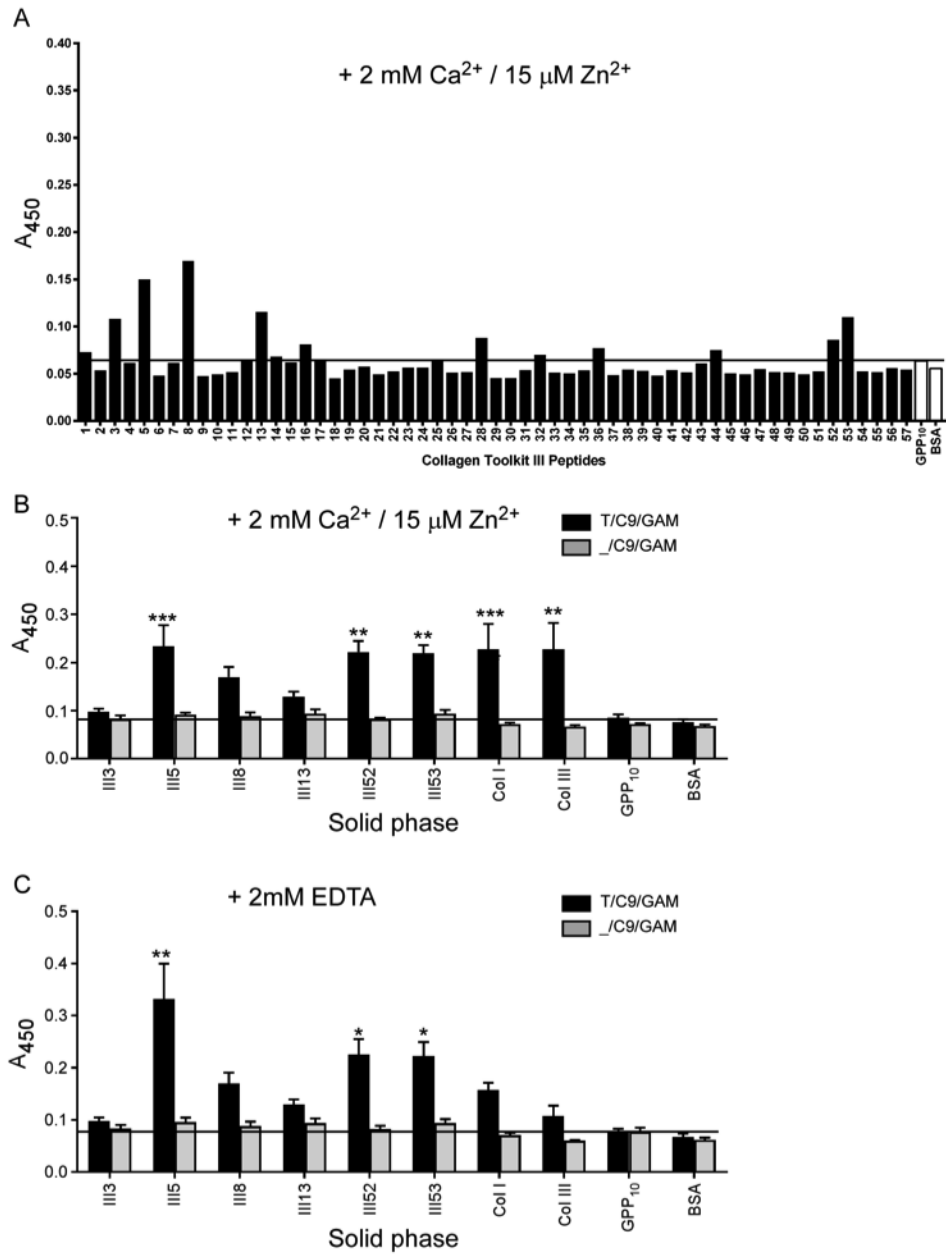


Fig. S1. Identification of specific TSP1-binding peptides within the triple-helical region of collagen III. (A) Binding of TSP1 to peptides from the Collagen Toolkit III in the presence of Ca²⁺ and Zn²⁺. The horizontal line represents the background binding of TSP1 to GPP₁₀. Each bar represents the mean of duplicate samples. (B) Binding of TSP1 to the indicated peptides from the Collagen Toolkit III, pepsin-digested collagen I (Col I), and pepsin-digested collagen III (Col III) in solid-phase binding assays. N=3 independent experiments. Each bar represents the mean, and the error bars indicate the s.e.m. (C) Binding of TSP1 to the indicated peptides in the presence of the Ca²⁺ chelator EDTA. N= 3 independent experiments. Each bar represents the mean, and the error bars indicate the s.e.m. In all panels, GPP₁₀ and BSA were included as negative controls. In B and C, pepsin-digested collagen I (Col I) and collagen III (Col III) were included as positive controls.

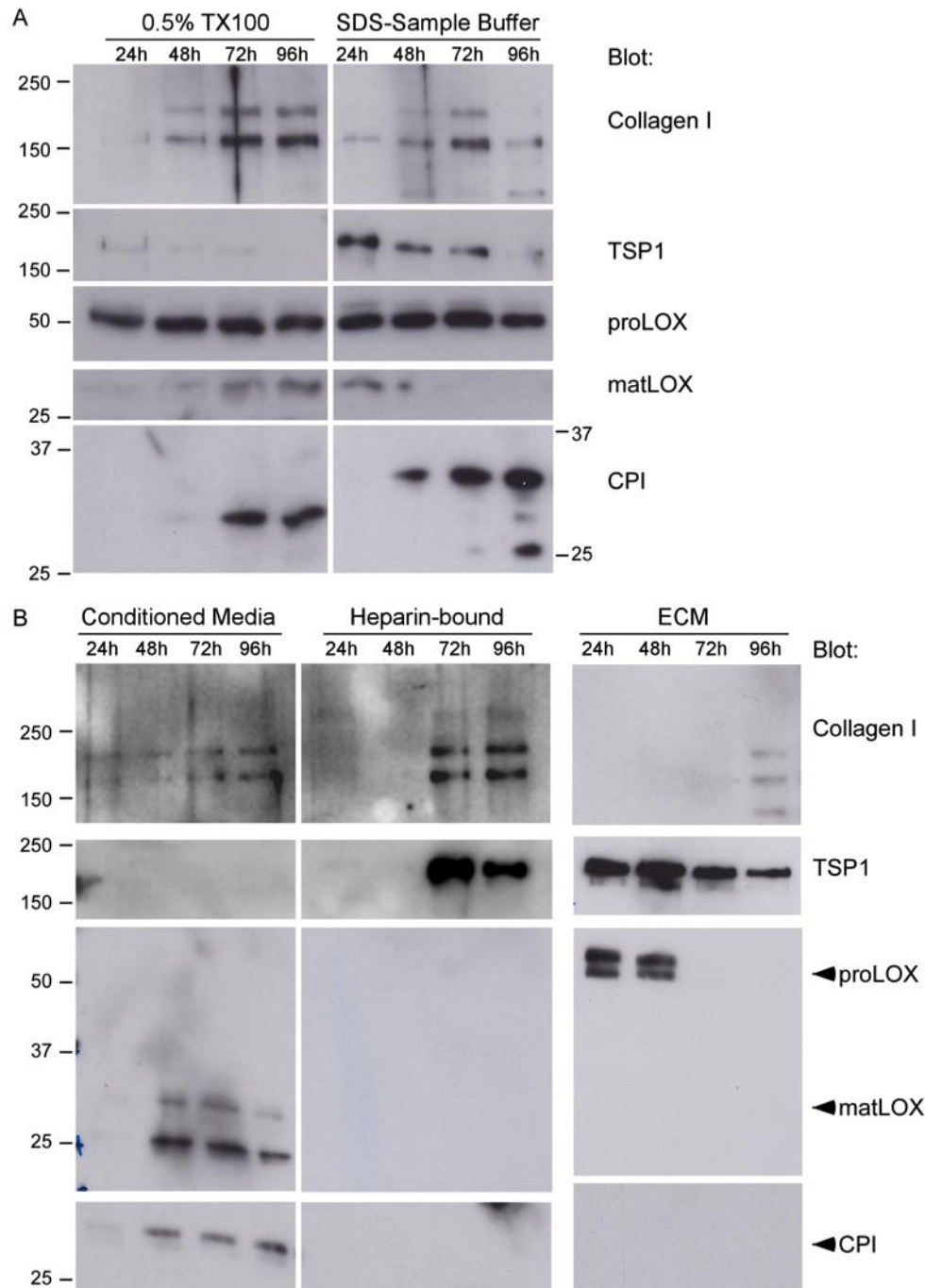


Fig. S2. Detection of collagen I, TSP1, FN, LOX and collagen I C-propeptide in cell fractions, CM, and ECM isolated from HDFs. Samples of human dermal fibroblasts (HDF) were prepared at the indicated timepoints, resolved on 10% SDS-PAGE gels under reducing conditions, and immunoblotted for the indicated proteins. **(A)** Cell lysates were prepared as indicated. In addition to the main CPI band of 30kDa, degradation products of CPI were observed in the sample buffer extracts at 96h. N = 2 independent experiments. **(B)** Conditioned media were analyzed directly (left panels) or after pull-down with heparin-agarose to concentrate heparin-binding proteins (center panels). ECM was isolated from the same cultures (right panels). The blot is representative of duplicate analysis. Non-contiguous lanes are indicated by gaps between panels. Molecular mass markers are indicated to the left of the blots in kDa. TX100, Triton X-100.

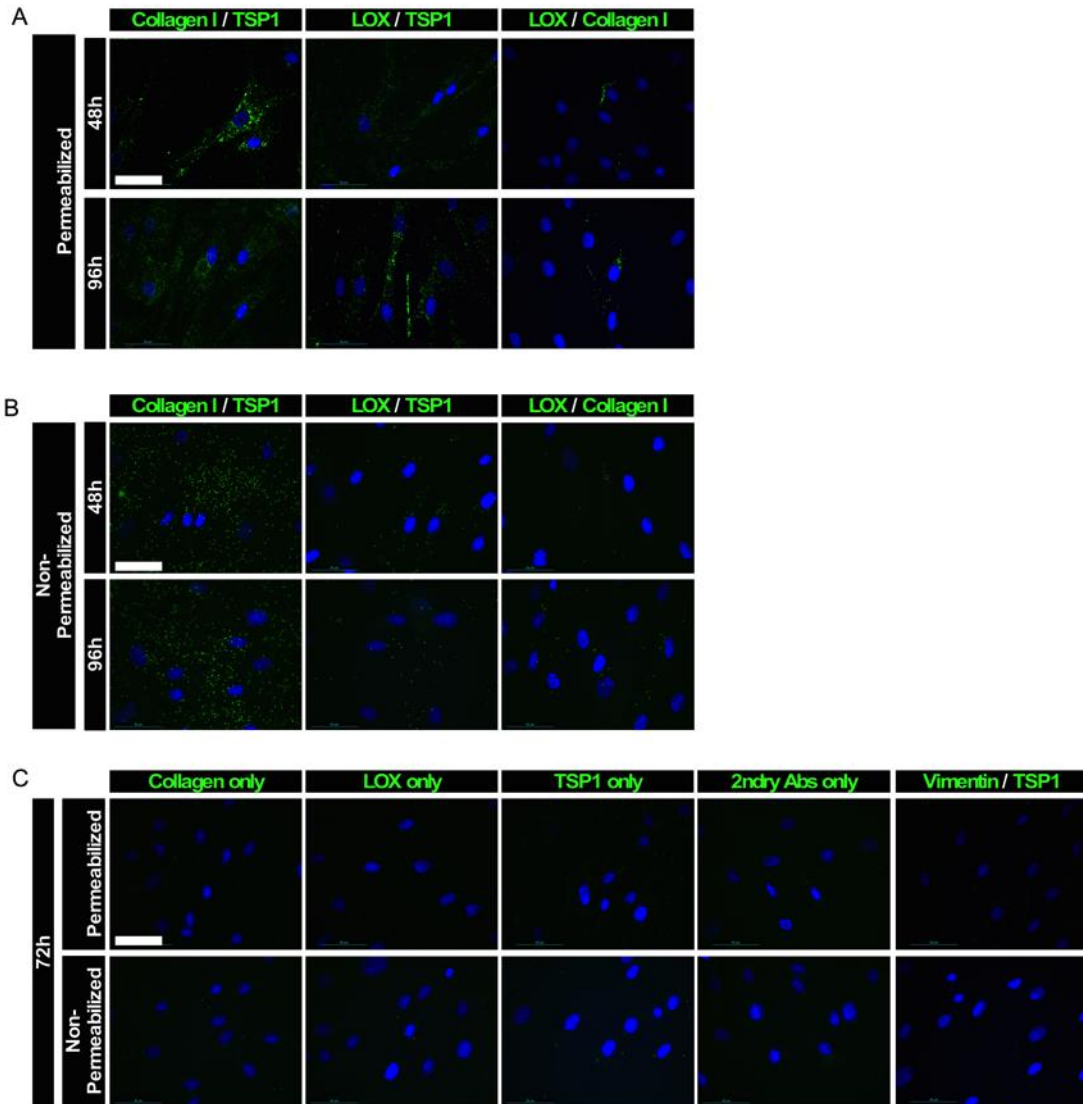


Fig. S3. Detection of the association of TSP1 with collagens I and II and LOX in HDFs by in situ proximity ligation. (A, B) In situ proximity ligation for detection of association between the indicated pairs of proteins in permeabilized (A) and non-permeabilized (B) HDF after culture for the indicated times. (C) Representative images of control in situ proximity ligation assays with single primary antibodies, secondary antibodies only, or vimentin and TSP1 as a pair of proteins predicted not to associate in HDF, analysed at 72h of culture. All images are representative of N = 4 independent experiments.

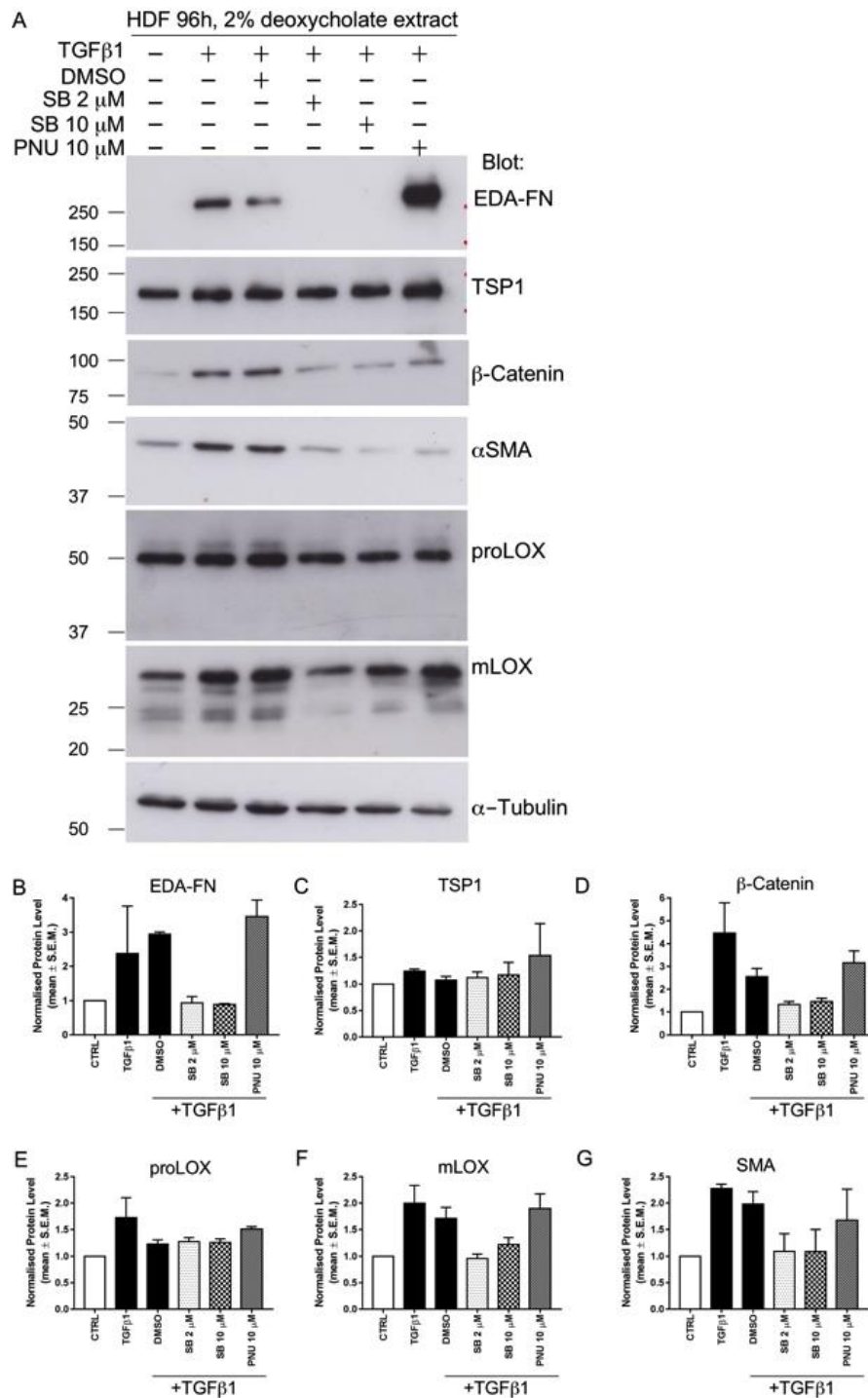


Fig. S4. TGF-β1-mediated induction of αSMA in HDFs. (A) HDFs were cultured in the presence or absence of TGF-β1, the TGF-βPR1 inhibitor SB-431542 (SB), or the β-catenin signaling inhibitor PNU-74654 (PNU) as indicated for 96h. Cell extracts were resolved on 10% polyacrylamide gels under reducing conditions and immunoblotted for the indicated proteins. The blot is representative of 3 experiments. Molecular mass markers are indicated to the left of the blot in kDa. (B-G) Quantification of EDA-FN (B), TSP1 (C), β-catenin (D), proLOX (E), mLOX (F), and αSMA (G) in the indicated treatment groups as normalized against α-tubulin. Each bar represents the mean from 3 independent experiments; error bars indicate s.e.m.

Table S1. Primary antibodies and the dilutions used in this study.

Antigen	Antibody Isotype	Supplier	Cat. Number #	WB	IF	In situ Proximity Ligation	ELISA
6-His tag	mouse monoclonal IgG2b	Abcam	ab18184				1:1,000
α -smooth muscle actin (SMA)	mouse monoclonal IgG2A	Sigma	A2547	1:1,000	1:1,000		
α -tubulin	rabbit polyclonal IgG	Abcam	ab4074	1:1,000			
Collagen α 1(I) C-propeptide (CPI)	rabbit polyclonal IgG	Larry Fisher, NIH	LF41	1:10,000			
Collagen I	mouse monoclonal IgG1	Abcam	ab 90395		1:1,500	1:1,500	
Collagen I	rabbit polyclonal IgG	Novus Biologicals	NB600-408	1:600	1:400	1:400	
EDA-Fibronectin	mouse monoclonal IgG1	Abcam	ab6328	1:1,000			
Lysyl oxidase (aa350 to the C-terminus)	rabbit monoclonal IgG	Abcam	ab174316 [EPR4025]	1:500	1:300	1:300	
Thrombospondin-1 (aa 25-48)	mouse monoclonal IgG1	SCBT	sc-393503	1:200	1:50	1:100	1:200
Vimentin	rabbit monoclonal IgG	Abcam	ab92547			1:1,000	

Key: ELISA = Enzyme-linked immunosorption assay; IF = immunofluorescence; Ig, immunoglobulin; WB = western blot.

Table S2. Secondary antibodies and the dilutions used in this study.

Antigen	Antibody Isotype and Conjugate	Supplier	Cat. Number #	WB	IF	In situ Proximity Ligation	ELISA
mouse IgG	MINUS PLA probe-conjugated donkey Ig	Sigma	DUO92004			1:5	
mouse IgG (Fab specific)	FITC-conjugated goat Ig	Sigma	F8771		1:500		
mouse IgG, IgA, IgM	HRP-conjugated goat IgG	Dako	P0447				1:15,000
mouse IgG, IgA, IgM	HRP-conjugated goat IgG	LI-COR	926-80010	1:100,000			
rabbit IgG	TRITC-conjugated sheep Ig	ThermoFisher Scientific	A16177		1:500		
rabbit IgG	PLUS PLA probe-conjugated donkey Ig	Sigma	DUO92002			1:5	
rabbit IgG, IgA, IgM	HRP-conjugated goat IgG	LI-COR	926-80011	1:100,000			

Key: ELISA = Enzyme-linked immunosorption assay; FITC, fluorescein isothiocyanate; HRP, horseradish peroxidase; IF = immunofluorescence; Ig, immunoglobulin; PLA, proximity ligation; WB = western blot.

Table S3. Chemicals used in this study.

Item	Supplier	Cat. Number #
Acetic Acid (HAc)	Sigma	A0808
Acetone	Fisher Scientific	10745863
Affi-gel® heparin beads	Bio-Rad	153-6173
Amersham ECL	GE Healthcare Life Sciences	RPN2209
Bromophenol Blue	Sigma	B0126
Calcium chloride (CaCl ₂)	Sigma	449709
Deoxycholic acid (DOCS)	Sigma	D2510
Detection Reagents Green	Sigma	DUO92014
DL-dithiothreitol (DTT)	Sigma	D0632
EDTA	Sigma	ED2SS
Gel Code Blue Stain reagent	ThermoFisher Scientific	24590
Glycerol	Sigma	G5516
Hydrochloric acid (HCl)	Sigma	H1758
L-ascorbic acid	Sigma	A4403
Leupeptin	Sigma	L2884
Methanol	Sigma	322415
N-Ethylmaleimide (NEM)	Sigma	E3876
Paraformaldehyde (PFA) 16% (w/v)	EMS or Alpha Aser	28906
Pepstatin A	Sigma	P5318
Pierce™ TMB Substrate Kit	ThermoFisher Scientific	34021
Sodium Dodecyl Sulfate (SDS)	Fisher Scientific	BP166-100
Triton-X100 (TX100)	Sigma	10789704001
Wash Buffer A	Sigma	DUO82047
Wash Buffer B	Sigma	DUO82048
Zinc sulfate heptahydrate (ZnSO ₄ · 7H ₂ O)	Sigma	Z0501

Table S4. Chemical inhibitors used in this study.

Target	Inhibitor	Supplier	Cat. Number #	CAS
Interaction between β -catenin and TCF (T cell factor)	PNU-74654	Sigma	P0052	113906-27-7
TGF- β RI Kinases (ALK4 and ALK5)	TGF- β RI Kinase Inhibitor VI, 616461	Calbiochem	SB431542	301836-41-9

Table S5. Proteins used in this study.

Protein	Supplier/Reference	Cat. Number #
Bovine serum albumin (BSA) Fraction V powder	GE Healthcare	K41-001
Recombinant human mini-pro-collagen I alpha 1 (rhProCOL1A1)	R&D	6220-CL-020
Human procollagen C-propeptide I (CPI)	Bourhis et al. 2012 Sharma et al. 2017	
Human procollagen C-propeptide III (CPIII)	Bourhis et al. 2012 Sharma et al. 2017	
Denatured collagen I (bovine)	Devro	NA
Native collagen I fibrils from equine tendon - Kollagen Reagens HORM Suspension	Takeda	NA
Pepsin-digested calf skin-derived collagen I	Sigma	C-3511
Pepsin-digested bovine tracheal cartilage-derived collagen II	Sigma	C-1188
Pepsin-digested human placenta-derived collagen III	Sigma	C-4407
Native purified human thrombospondin-1 (TSP1)	Cell Sciences Inc	CSI19832B
Recombinant human C-terminal MYC/DDK-tagged lysyl oxidase (proLOX)	Origene	TP313323
Recombinant human TGF β 1	R&D	240-B

Table S6. Buffers used in this study.

Buffer	Composition	Experiment
Protease inhibitor cocktail	2 mM NEM, 10 mM leupeptin, 20 mM pepstatin A	Collagen extraction from mouse skin
Incubation Buffer (IB)	Tris-Buffered Saline (TBS), 1 mg/ml BSA, 0.1 % (v/v) Tween-20	Solid phase binding assay
SDS-PAGE sample buffer	100 mM Tris.HCl, pH 6.8, 4 % SDS, 0.2 % bromophenol blue, 20 % glycerol containing 100 mM DTT	WB
DOCS-buffer	2 % deoxycholic acid, 20 mM Tris pH 8.8, 2 mM N-ethylmaleimide	WB
Immunoblot Blocking Buffer (BB)	TBS containing 2 % (w/v) semi-skimmed dried milk and 0.2 % (v/v) Tween 20	WB

Key: WB = western blot.